

Cell isolation with Strep-Tactin® TACS Agarose columns (1 ml)

for whole blood, buffy coat, PBMCs or other single cell suspensions

1. Required material

Cat. No.	Product	Required/isolation
6-6310-001	Strep-Tactin® TACS Agarose Column, 1 ml	1
-	Protein of choice fused to a Strep-tag®II or Twin-Strep-tag®	50 µg*
6-6325-001	Biotin stock solution, 100 mM, 1 ml	200 µl
-	Buffer for cell isolation, e.g. 1x PBS containing 1 mM EDTA and 0.5% BSA	~ 70-80 ml
6-6331-001	TACS Column Adapter (1 ml column)	1

*Amounts refer to a 50 kDa Twin-Strep-tag® protein

2. Initial preparations

2.1. Reagent preparation

Allow the reagents to equilibrate to **room temperature** (RT) prior to use. For a sterile isolation, work under a safety cabinet. **The following volumes will be sufficient for one selection process.**



Watch this How-to video to see an exemplary isolation:

https://www.youtube.com/watch?v=OPL_-uNjFZQ



- 2.1.1.** Prepare a cell isolation buffer of choice, e.g. 1x PBS containing 1 mM EDTA and 0.5% BSA. The buffer should not contain biotin. Degas buffer before use, as air bubbles could block the column.
- 2.1.2.** Dilute your Strep-tag®II or Twin-Strep-tag® fusion protein to a concentration of **50 µg/ml** with cell isolation buffer.



Titration of optimal isolation conditions might be necessary. The following instructions are an example for isolating cells with a **50 kDa** protein fused to a Twin-Strep-tag®.

- 2.1.3.** Prepare **1 mM** Biotin Elution Buffer by diluting **200 µl** of 100 mM Biotin stock solution in **20 ml** cell isolation buffer. Mix thoroughly.

2.2. Sample preparation

- 2.2.1.** For **PBMCs** or other single cell suspensions: Prepare cells and resuspend up to **3 x 10⁸ cells/ 5 ml** cell isolation buffer.
- 2.2.2.** For **whole blood or buffy coat**: Dilute in a 3:1 ratio with cell isolation buffer, e.g. dilute **9 ml** whole blood with **3 ml** cell isolation buffer. Mix gently by pipetting up and down. To remove clumps and to prevent aggregates, pass sample through a 40 µm nylon mesh before separation.

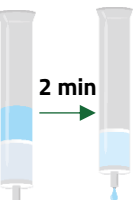
2.3. Column preparation



2.3.1. Remove the cap and cut the sealed end of the column at notch. Allow the storage solution to drain. Place the Strep-Tactin® TACS Agarose Column into the TACS Column Adapter.



2.3.2. Wash the Strep-Tactin® TACS Agarose Column by applying **5 ml** cell isolation buffer and allow the buffer solution to enter the packed bed completely.



2.3.3. Load 1 ml Twin-Strep-tag® fusion protein (2.1.2.) onto the Strep-Tactin® TACS Agarose Column. Let the protein solution enter the packed bed completely. Incubate for **2 min**.



2.3.4. Wash the Strep-Tactin® TACS Agarose Column with **2 ml** cell isolation buffer. Discard flow-through and change collection tube. The Strep-Tactin® TACS Agarose Column is now ready for cell isolation.

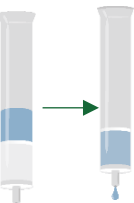


Do not interrupt the procedure for **more than 60 min**.

If you plan to isolate your cells from **PBMCs** or other single cell suspensions follow chapter **3.1**. For isolation from **whole blood or buffy coat** follow chapter **3.2**.

3. Protocol

3.1. Cell isolation from PBMCs or other single cell suspensions

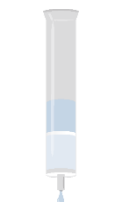


3.1.1. Load

Apply cell suspension (2.2.1.) in steps of max. **5 ml**. Collect flow-through containing unlabeled cells.



If you expect more than 5×10^7 target cells you can apply the flow-through a second time to maximize the yield.



3.1.2. Wash

Apply **4 x 10 ml** cell isolation buffer. (In each step: Let the buffer solution enter the gel bed completely).

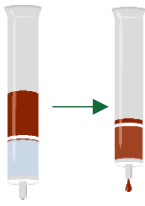


3.1.3. Elute

From this step on your flow-through contains your target cells. Use a new collection tube. Apply **1 ml** Biotin Elution Buffer (2.1.3.) and incubate for **5 min**. Elute target cells by applying **9 ml** Biotin Elution Buffer. Elute a second time with additional **10 ml** Biotin Elution Buffer.

3.1.4. Optional: Apply additional **5 ml** of cell isolation buffer to the column and immediately centrifuge at **310 x g** for **2 min** to increase yield.

3.2. Cell isolation from whole blood or buffy coat



3.2.1. Load

Apply diluted whole blood or buffy coat (2.2.2.) in steps of max. **5 ml**. Collect flow-through containing unlabeled cells.



3.2.2. Wash

Apply **4x 10 ml** cell isolation buffer. (In each step: Let the buffer solution enter the gel bed completely).



3.2.3. Elute

From this step on your flow-through contains your target cells. Use a new collection tube. Apply **1 ml** Biotin Elution Buffer (2.1.3.) and incubate for **5 min**. Elute target cells by applying **9 ml** Biotin Elution Buffer. Elute a second time with additional **10 ml** Biotin Elution Buffer.

3.2.4. Optional for buffy coat: Apply additional **5 ml** of cell isolation buffer to the column and immediately centrifuge at **310 x g** for **2 min** to increase yield.

3.3. Further procedure

Centrifuge your eluted cell suspension for **10 min** at **300 x g**. Discard the supernatant and dissolve cell pellet in your desired buffer.



If you plan to continue with a biotin-sensitive assay, please remove biotin by washing with **50 ml** cell isolation buffer twice. Discard supernatant completely.

4. Troubleshooting

Low yield

Option 1

Check for biotin contamination in your samples.

Option 2

Re-apply flow-through (depleted sample) to the column.

Low purity

Invert columns after each wash step three times.

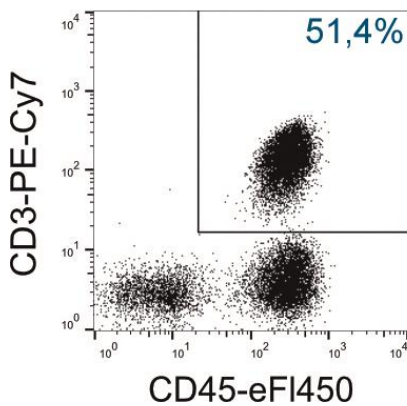
5. Example data

5.1. PBMCs

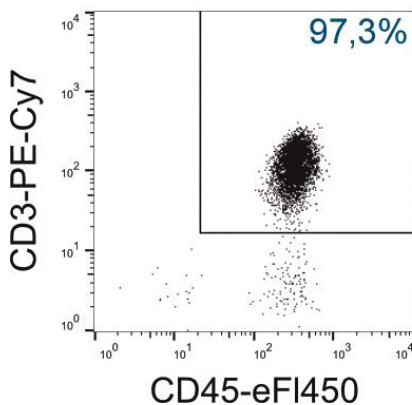
Separation of CD3⁺ T cells from 5 ml PBMCs (containing 3 x 10⁸ cells) using a Fab Fragment against CD3 fused to a Twin-Strep-tag®. Unlysed cells were stained with CD3-PE-Cy7 (OKT-3) / CD45-eFl450 (2D1) and analyzed by flow cytometry (CyAn ADP, BC). Dead cells were excluded from the analysis using PI staining. Doublet and debris discrimination were performed using different FSC/SSC signals.

PBMCs

Before isolation



After isolation

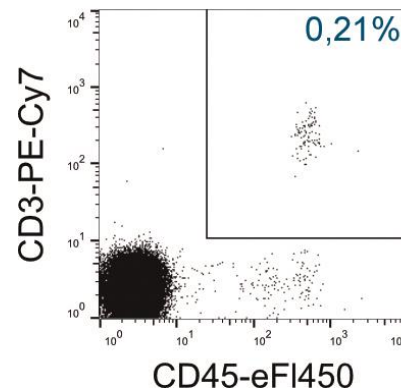


5.2. Buffy coat

Separation of CD3⁺ T cells from buffy coat sample using a Fab Fragment against CD3 fused to a Twin-Strep-tag®. Unlysed cells were stained with CD3-PE-Cy7 (OKT-3) / CD45-eFl450 (2D1) and analyzed by flow cytometry (CyAn ADP, BC). Dead cells were excluded from the analysis using PI staining. Doublet and debris discrimination were performed using different FSC/SSC signals.

Buffy coat

Before isolation



After isolation

